

INHIBITION OF CO₂ FIXATION IN INTACT SPINACH CHLOROPLASTS
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SUMMARY: Glycerate-3-P inhibits CO₂ fixation of isolated spinach chloroplasts at concentrations higher than 1 mM but does not inhibit O₂ evolution. Glycerate-3-P inhibition of photosynthesis is not overcome by higher bicarbonate concentrations.

INTRODUCTION.

Intermediates of the photosynthetic carbon reduction cycle such as ribose-5-P, fructose-1,6-diP, glyceraldehyde-3-P, dihydroxyacetone-P and 3-phosphoglyceric acid (3-PGA) can stimulate photosynthesis and overcome the initial lag phase of photosynthesis when isolated chloroplasts are assimilating CO₂ in the light (1). It has been proposed that the stimulation of CO₂ fixation and also the overcoming of the lag phase for CO₂ fixation in isolated chloroplasts by the addition of intermediates to the reaction media were due to a filling of depleted substrate pools. (2). However, 3-PGA appears to have a unique two-phase effect on chloroplast activities because we have observed that 1 to 2 mM concentrations of 3-PGA can also inhibit CO₂ fixation.

In this paper we report on the effects of exogenously added 3-PGA on the rates of CO₂ fixation and O₂ evolution of isolated chloroplast suspensions fixing CO₂ in the light in order to elucidate the site of 3-PGA regulation.

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MATERIALS AND METHODS.

Chloroplasts were isolated from spinach leaves (*Spinacia oleracea* L.) by the following procedure: 10 g of diced leaves from the growth chamber were homogenized for 2 sec in 40 ml of a solution of pH 6.8, containing 0.33 M sorbitol, 50 mM Hepes-KOH buffer, 2 mM EDTA, 1 mM $\text{Na}_4\text{P}_2\text{O}_7$, 1 mM MgCl_2 and 1 mM MnCl_2 . The homogenate was filtered through Miracloth, centrifuged at $755 \times g$ for 90 sec., and the pellet of chloroplasts was suspended in 2 ml of the above solution containing 5 mM ascorbate at pH 7.8. The chlorophyll concentration was usually between 60-100 μg chlorophyll ml^{-1} .

RESULTS.

The data in Table 1 compare the rates of CO_2 fixation in isolated chloroplasts treated with 0 and 2 mM 3-PGA. The reactions were in photosynthetic reaction systems containing 1, 5, and 10 mM bicarbonate. Strong inhibition of CO_2 fixation was always observed with 2 mM 3-PGA at all bicarbonate concentrations.

Data for the effects of 3-PGA on O_2 evolution from isolated chloroplasts fixing CO_2 in the reaction mixtures are also listed in Table 1. The treatments consisted of 0 and 2 mM 3-PGA at 1, 5, 10 mM bicarbonate levels. Clearly, 3-PGA did not inhibit O_2 evolution in chloroplasts fixing CO_2 .

DISCUSSION.

The experimental evidences presented in Table 1 show that 3-PGA inhibited CO_2 fixation but not O_2 evolution when intact chloroplasts were assimilating CO_2 . We assume therefore that the site for 3-PGA stimulated inhibition of CO_2 fixation was not associated with the photosystem, the electron transport system nor with the reductive phase of carbon metabolism. If the reductive step of the Calvin cycle or any of the pathways involved in the transfer of

Table 1

THE EFFECTS OF 3-PHOSPHOGLYCERATE ON CO₂ FIXATION AND O₂ EVOLUTION OF INTACT SPINACH CHLOROPLASTS*

Concentration of 3-PGA	Concentration of HCO ₃ ⁻ in Each Reaction		
	1 mM	5 mM	10 mM
	μmoles CO ₂ fixed/mg chlorophyll ⁻¹ hour ⁻¹		
Control	14.89	22.67	23.49
2 mM	5.89	7.76	9.35
	μmoles of O ₂ evolved/mg chlorophyll ⁻¹ hour ⁻¹		
Control	9.85	16.10	22.17
2 mM	14.09	17.11	26.01

*The rate of CO₂ fixation was determined by ¹⁴CO₂ (25 μc μmole⁻¹) uptake in test tubes at 25° C in a water bath. Illumination was by four 150 w flood lamps at 120 V. Oxygen evolution was measured polarographically with a modified Clark-type electrode inserted into a thermostated, 1 ml, lucite chamber at 25° C. Light was provided by a 150 w flood lamp with a 5 cm water filter. Light intensities at the reaction vessels were comparable in the CO₂ and O₂ experiments. The reaction mixtures contained 0.33 M sorbitol, 50 mM Hepes KOH, 1 mM MgCl₂, 1 mM MnCl₂, 2 mM EDTA, 1 mM Na₄P₂O₇, 5 mM ascorbate. In addition the mixtures contained 3-phosphoglycerate and NaHCO₃ in the concentrations listed above.

reducing power from light to the Calvin cycle were inhibited by the presence of 3-PGA this should be reflected as a reduction of the rate of O₂ evolution from intact chloroplasts in the process of photosynthesis. It appears therefore that the site of the 3-PGA-stimulated inhibition of CO₂ fixation is associated with processes which regenerate the phosphorylated carbohydrate acceptor for CO₂. We consider that in our preparations the carboxylation step is not involved as the site for the 3-PGA inhibition although Paulsen and Lane (4) showed that 3-PGA inhibited purified ribulose-1,5-diP carboxylase. They

observed that 3-PGA performed as a competitive inhibitor for bicarbonate. Our studies clearly show that the 3-PGA inhibition of CO₂ fixation of isolated plastids was not affected by higher amounts of bicarbonate.

On the other hand, Heldt (3) has demonstrated in spinach chloroplasts that 3-PGA can be exchanged for glyceraldehyde-3-P. This exchange would result in an inhibition of CO₂ fixation but not in O₂ evolution. We believe that this mechanism would explain our observations. Finally, it is possible that in the presence of excess 3-PGA, glycerate-1,3-diP accumulates which may inhibit a reaction of the photosynthetic carbon reduction cycle.

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